



PRODUCTION OF REFERENCE MATERIALS AND HOMOGENEITY AND STABILITY TESTING

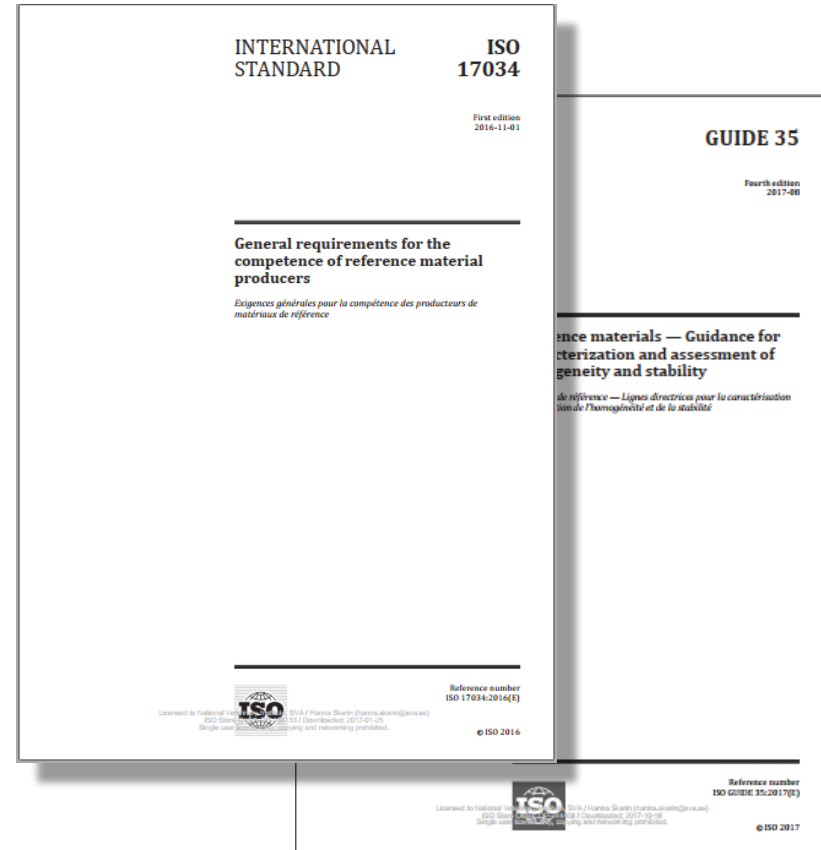
Sevinc Ferrari

EURL-Campylobacter Training Course 9 October 2019

REQUIREMENTS AND GUIDANCE FOR PRODUCTION OF REFERENCE MATERIALS

There is an International Standard and three ISO Guides that support production and certification of reference materials.

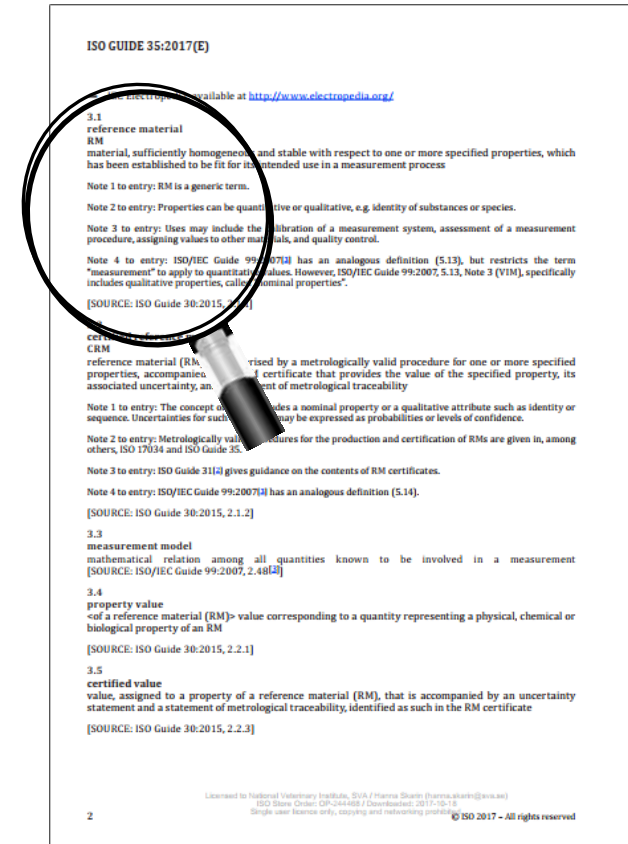
- ISO 17034 - general requirements
- ISO Guide 35 - guidance on technical issues, assessment of homogeneity and stability etc.
- ISO Guide 31 - contents of certificates for certified RMs
- ISO Guide 30 - terms and definitions



DEFINITION

Reference material (RM) is defined in ISO 17034 and Guide 35 as

“material, sufficiently homogeneous and stable with respect to one or more specified properties, which has been established to be fit for its intended use in a measurement process”



FOLLOWING REQUIREMENTS MUST BE FULFILLED FOR RMS

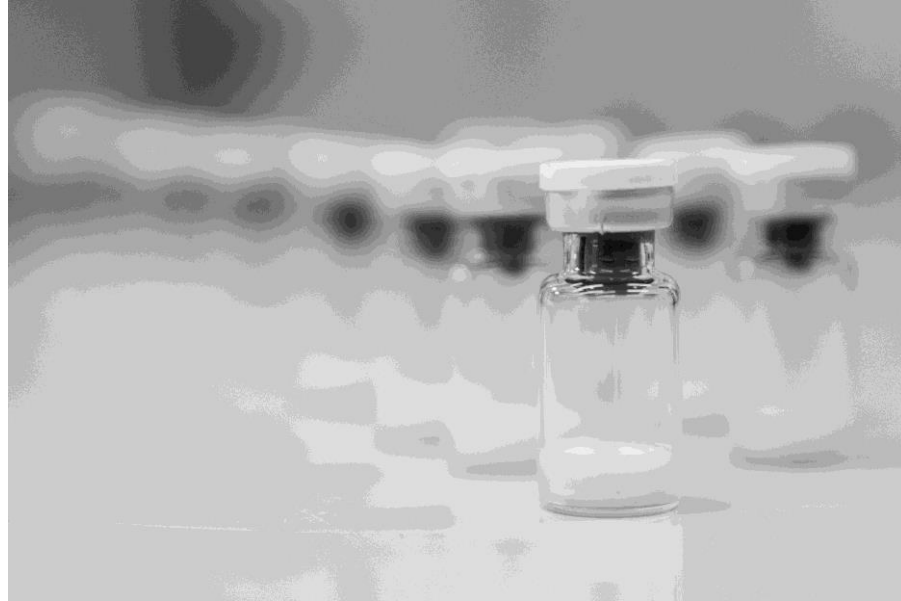
- representative for its intended use
representative = resemble routine samples as much as possible
- homogeneity specified within defined limits
- stability specified within limits over a specified period of time

***FOR CAMPYLOBACTER THERE ARE TWO PRODUCERS OF
QUANTITATIVE MICROBIOLOGICAL RMS***

- National Food Agency (Sweden) - Freeze-dried material
- Biosisto (The Netherlands) - Cryo cultures



LYOPHILIZATION / FREEZE-DRYING



A CLOSER LOOK AT THE FREEZE-DRY PROCESS

- **Freezing** - formation of ice crystals
- **Primary drying** - removal of ice crystals
Sublimation (at -35°C)
- **Secondary drying** - removal of unfrozen water
Desorption (at 25°C)

FREEZE-DRYING CHALLENGES

- Anaerobic and micro-aerophilic organisms, such as *Campylobacter* spp., are more difficult to freeze-dry due to their sensitivity to oxygen
- Gram negative bacteria show lower freeze-drying survival than gram positive
- A successful freeze-drying process doesn't guarantee a long shelf-life

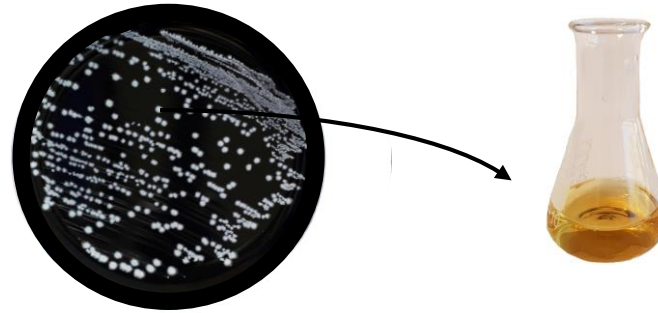
Temperature and humidity are important factors that affect the storage stability.

FREEZE-DRYING PROCEDURE

- Selection and identification of the test strain
- Decide the batch size
- Culture
- Formulation
- Filling vials
- Partial stoppering
- Load freeze-dryer
- Freeze-drying
- Cap/seal

CULTURE

Enrichment in 10 ml BHI



Inoculate a small part of a colony rather than a whole colony.

Incubation time differ for different strains and species

- *C. jejuni* 18h
- *C. coli* 18h
- *C. lari* 24h
- *C. hyointestinalis* 50h
- *C. lanienae* 48h
- *C. upsaliensis* 48h
- *C. helveticus* 22h



FORMULATION



1

Culture

Peptone saline water

Vortex and dilution

2



culture + SPG

3



SPG

ISB on ice with magnetic stirrer

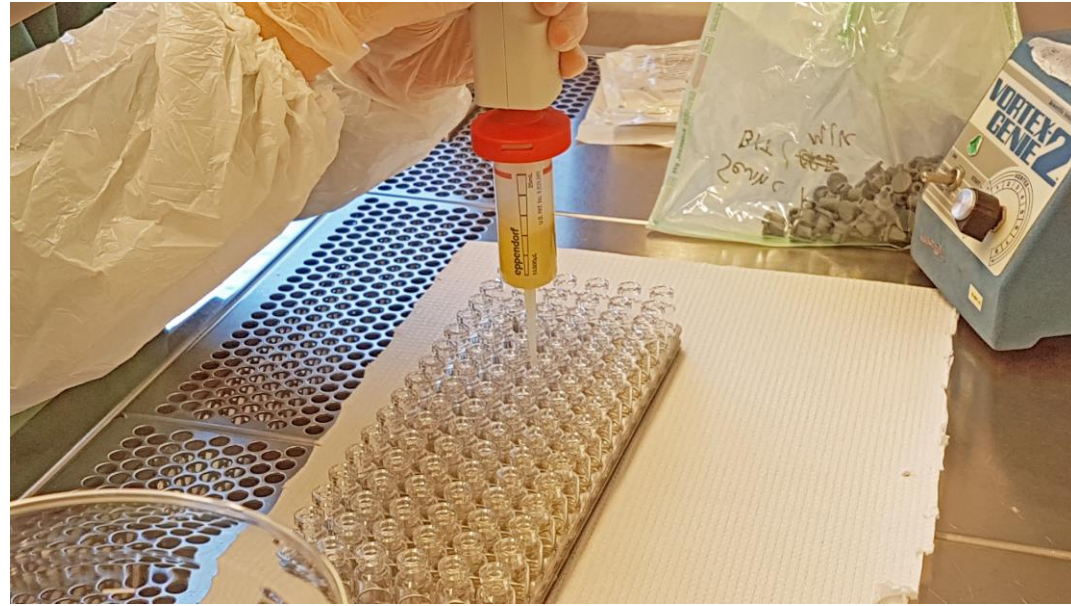
Protective agents

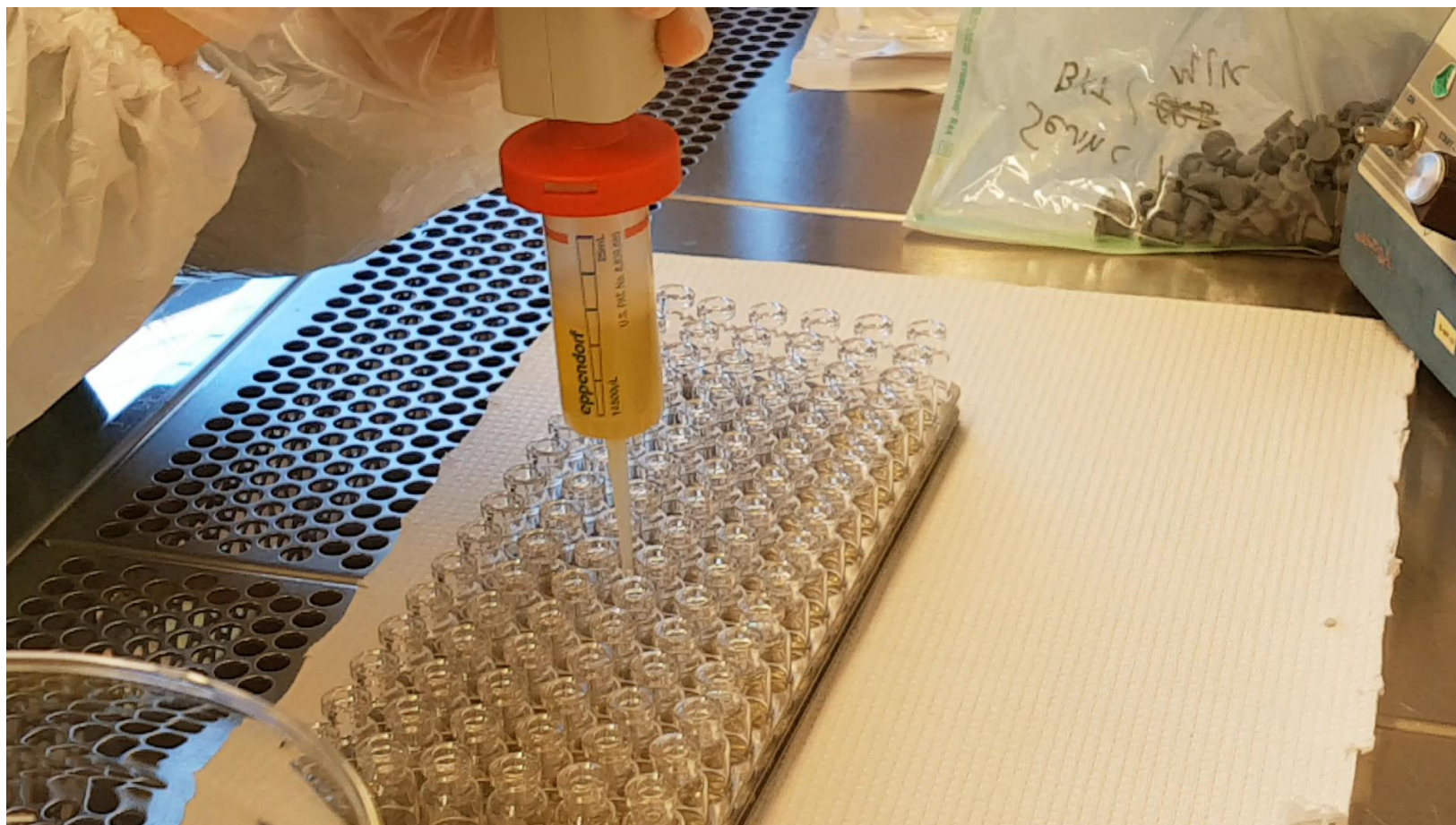
SPG: saccharose phosphate glutamate, and peptone

ISB: horse serum, inositol, nutrient broth

culture + SPG to ISB

FILLING VIALS





PARTIAL STOPPERING



Partially stoppered vial

LOAD FREEZE-DRYER AND FREEZE-DRY



CAP/SEAL



THE END PRODUCT

Dried cake



Uniform colour and texture of
the cake after freeze-drying

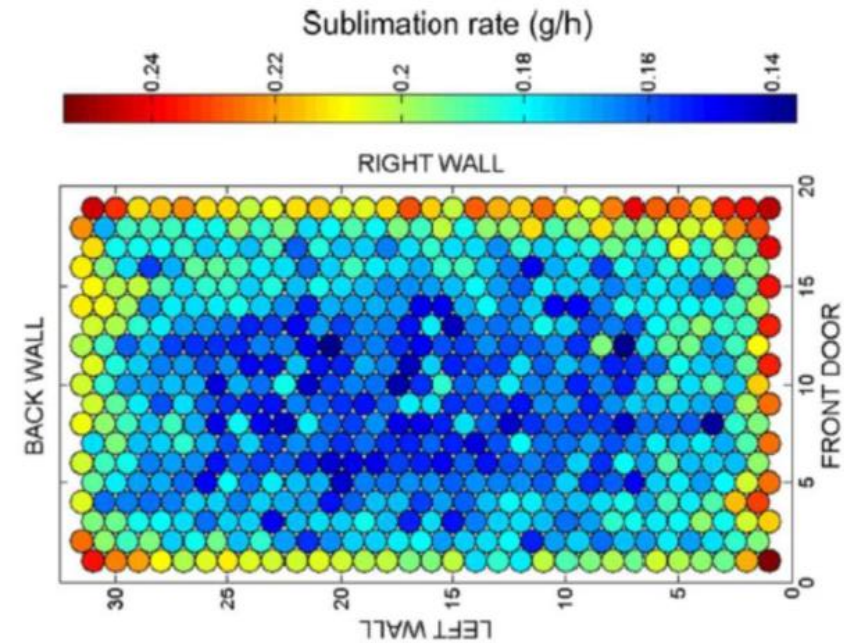
LIMITATIONS OF BATCH FREEZE-DRYING

Variability in sublimation rate

Uneven heat transfer



Vial-to-vial product variability



Kauppinen et al.

HOMOGENEITY ASSESSMENT

An experimental homogeneity study is provided in ISO Guide 35

“experimental homogeneity tests require measurements of a representative number of randomly chosen units. The units can be chosen for example by random selection, stratified random selection or systematic selection from a random start point.”

HOMOGENEITY STUDY FOR QUANTITATIVE PROPERTIES

N_{prod} = the total number of units produced

N_{min} = recommended minimum number of units

$$N_{min} = \max(10, \sqrt[3]{N_{prod}})$$

Example 1: You prepare 3 000 vials of a RM and intend to undertake a homogeneity study. The cube root of 3000 is 14,4. This study requires 15 vials for the homogeneity study.

Example 2. You prepare 500 vials of a RM and intend to undertake a homogeneity study. The cube root of 500 is 7,9. This study requires 10 vials for the homogeneity study.

HOMOGENEITY STUDY ON SMALL BATCHES

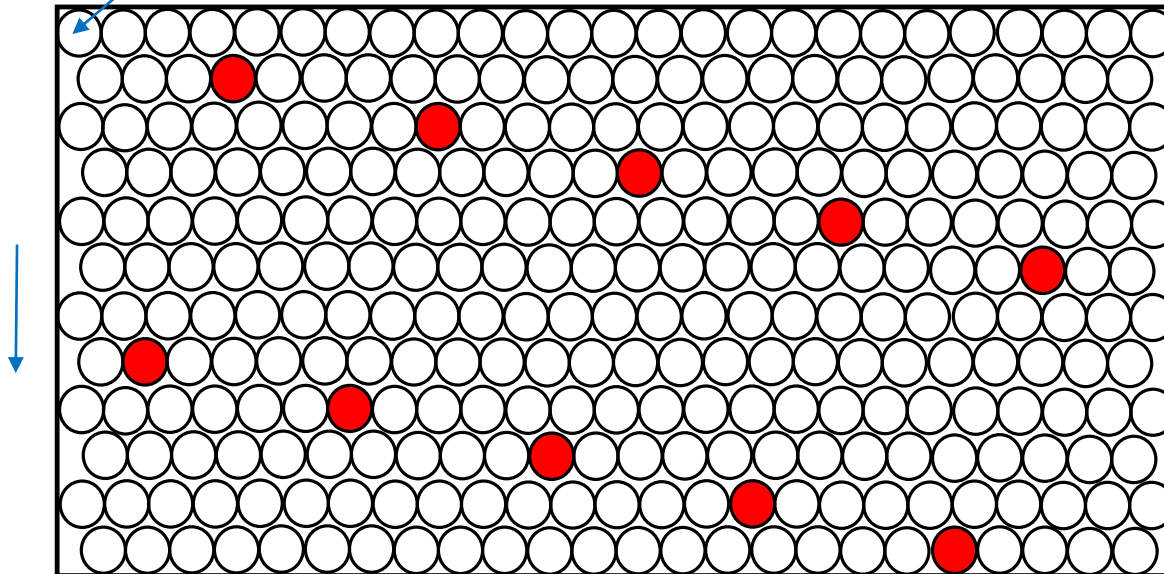
$N_{prod} < 100$ units

homogeneity should be assessed on the larger of three units or 10% of the batch size

SYSTEMATIC SAMPLING FOR THE HOMOGENEITY STUDY

$N_{prod} = 294$ vials

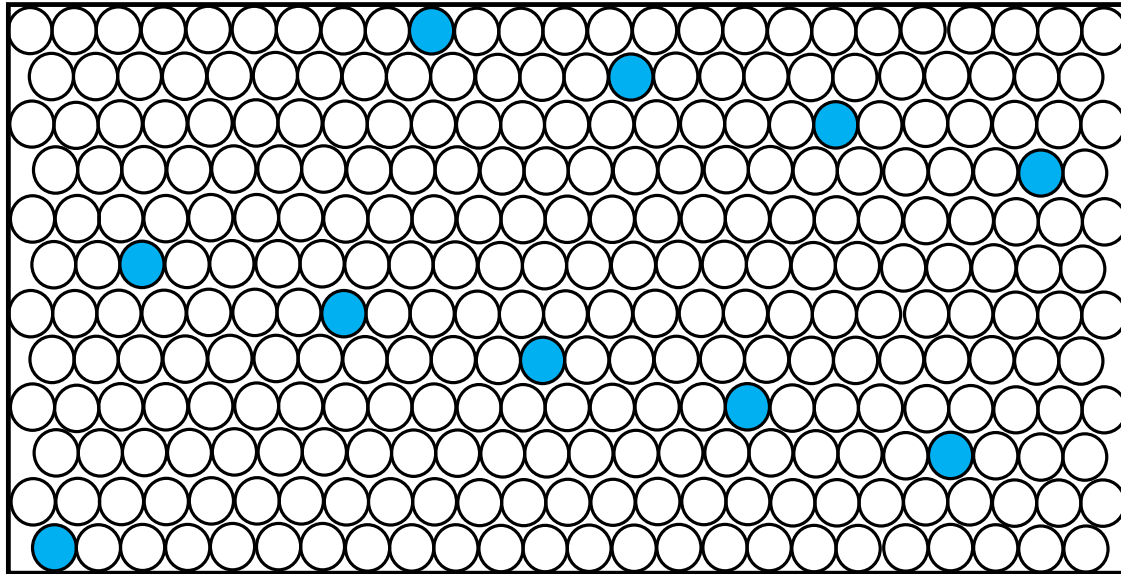
Production start point



Important to have an interval between the samples in order to cover the whole filling line.

In this example every 29th vial is selected for the homogeneity study.

Systematic selection from a random start point



STATISTICAL TREATMENT

Number of randomly chosen units = 10

Homogeneity: $s < 0.15 \log \text{CFU}$, $\text{max-min} < 0.5 \log \text{CFU}$

An example

C. jejuni (SVA021)

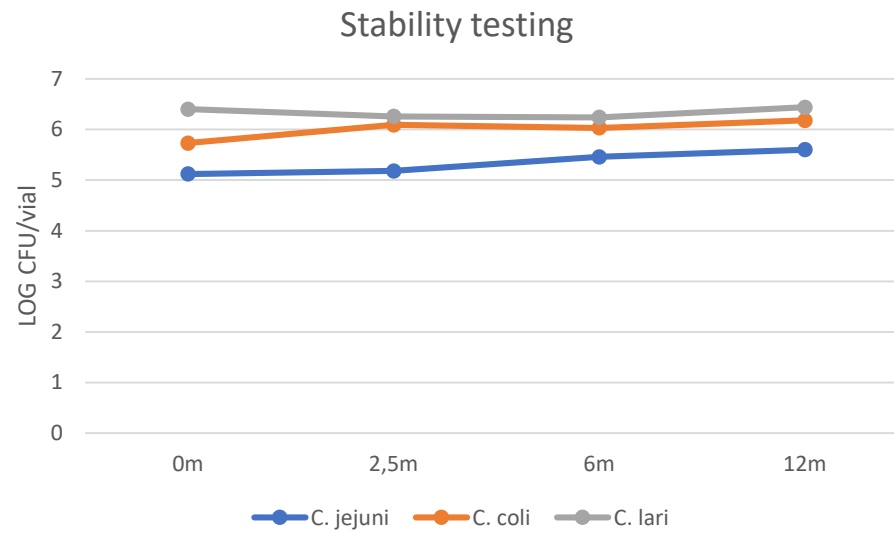
Sample	Plate 1 (0 dilution) No of CFU	log CFU/vial
1	187	4,27
2	189,5	4,28
3	187,5	4,27
4	190	4,28
5	188	4,27
6	158	4,20
7	164,5	4,22
8	212	4,33
9	194,5	4,29
10	233	4,37

	CFU/ampoule	log CFU/ampoule
Mv	190	4,28
s	21	0,05
Min	158	4,20
Max	233	4,37
Max-Min	75	0,17

STABILITY MONITORING

- Most RMs are stored for extended periods
 - necessary to assess the stability during storage conditions
- Nearly all RMs have to be transported to the location of use
 - necessary to assess the stability during transport conditions

STABILITY MONITORING: *C. JEJUNI*, *C. COLI* AND *C. LARI*



TESTING OF PT

The whole PT is tested

- when designing the PT (pilot tests)
- right before sending the PT (pre-PT)
- when most labs have received their PT (2 days after sending)
- the last date set for running the PT



Thank you for your attention!

Questions?

