

# RESULTS OF PROFICIENCY TESTS NO. 23 AND 24



Helena Höök
EURL-*Campylobacter*Workshop 2019





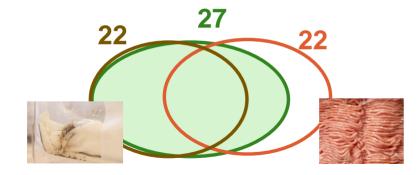




Thank you for your participation and for providing information in the questback reports!

### **NUMBERS OF PARTICIPANTS**

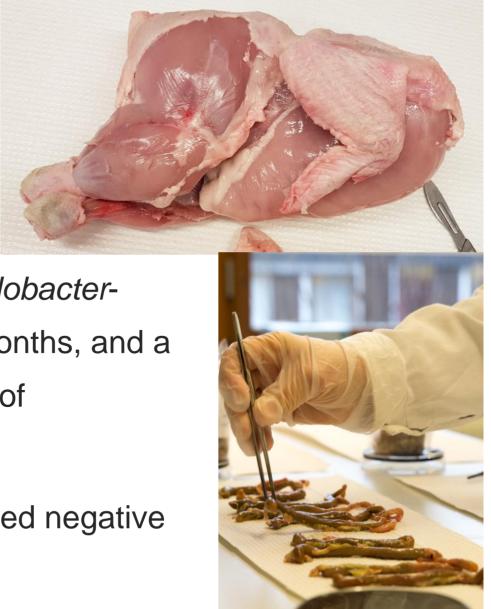
Year	2019	2018	2017	2016	2015	2014	2013	2012	2011
	PT 23	PT 21	PT 19	PT 17	PT 15	PT 13	PT 11	PT 9	PT 8
Enumeration	35	37	36	36	36	35	36	33	33
	PT 24	PT 22	PT 20	PT 18	PT 16	PT 14	PT 12	PT 9	PT 8
Detection & species id	33	31	34	33	32	36	34	36	34



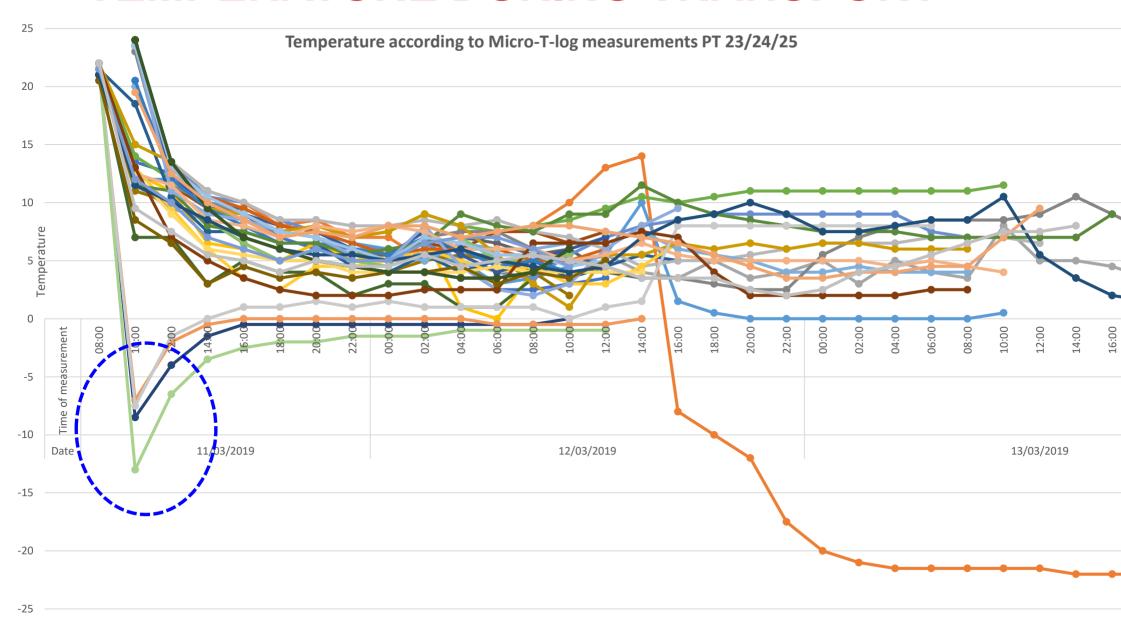


### **CAMPYLOBACTER-FREE MATRICES**

- Chicken meat (PT 23, PT 24)
- Caecal material (PT 24)
- Litter material (PT 24)
- All from a producer with no Campylobacterpositive broiler flocks for several months, and a slaughterhouse with very low level of Campylobacter-positive flocks
- Meat, litter and ceacal material tested negative for presence of Campylobacter



### **TEMPERATURE DURING TRANSPORT**





### **PROFICIENCY TEST NO. 23**

The objective was to assess the performance of the NRLs to enumerate (and voluntary species identify) *Campylobacter* in chicken meat.

- Enumeration and confirmation of Campylobacter spp. in chicken meat
- Species identification of Campylobacter (voluntary)
- Recommended method ISO 10272:2017, but other methods allowed
- Should allow enumeration of between 10 and 10<sup>5</sup> cfu
   Campylobacter/g chicken meat



### PT 23: CONTENTS AND PROCEDURE

- Chicken meat (about 120 g) to be divided into 10 portions of 10 g
- 10 vials with freeze-dried sample (with or without Campylobacter)
- Homogenise and make a initial dilution of 10<sup>-1</sup>
- Follow the method(s) of choice for
  - enumeration
  - species identification (voluntary)



of Campylobacter spp.

### **DESCRIPTION OF THE 10 VIALS IN PT 23**

Sample No.	Species	Level (log cfu/vial)	Batch No.
1	C. jejuni	3.71	SLV306
2	C. lari	4.82	SLV248
3	Negative		SLV289
4	Escherichia coli	4.46	SLV150
5	C. lari	4.04	SLV299
6	C. jejuni	3.71	SLV306
7	C. jejuni + Escherichia coli	3.50 4.00	SLV313
8	C. coli	5.67	SLV287
9	C. jejuni	4.47	SVA010
10	C. coli	5.67	SLV287

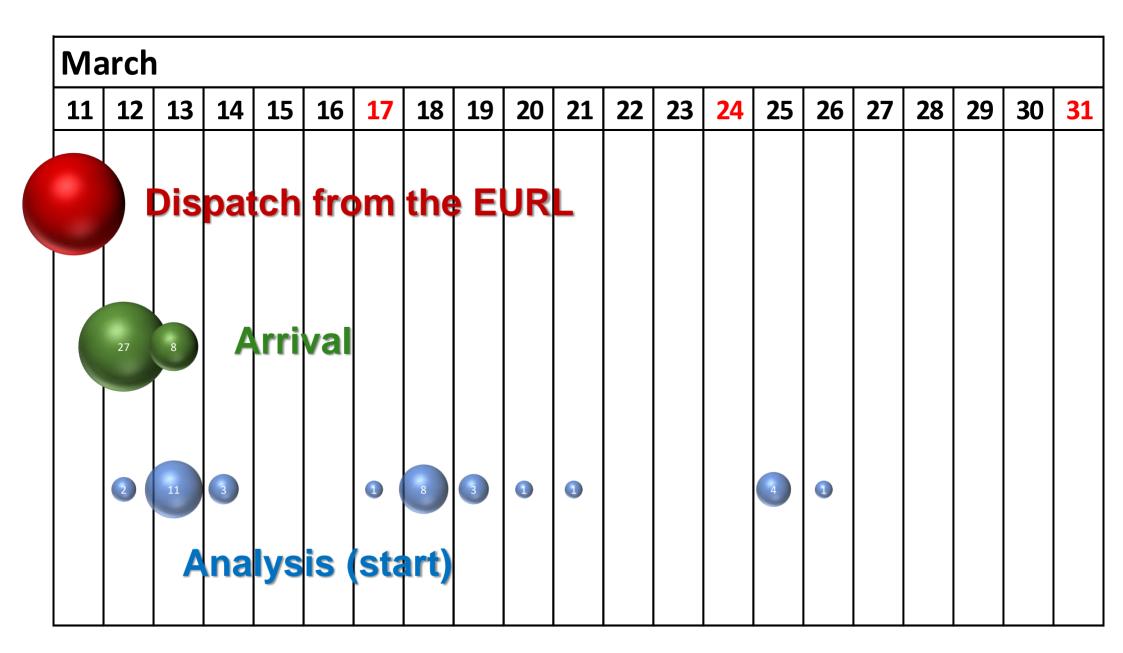
## PT 23: QUALITY CONTROL

- Vials produced by the National Food Agency
- Vials tested for homogeneity and stability by the producer and in triplicates by EURL
- Enumerations with chicken meat for control of *Campylobacter* levels and homogeneity
- Tested three times, once before and twice after dispatch





#### PT 23: TIME TO ARRIVAL & START OF ANALYSIS



# PT 23: METHODS

Reported method for enumeration	No. of NRLs
ISO 10272:2017	32
NMKL 119, 3rd ed. 2007	2
Intern method	1





## WHAT'S IN THE RESULTS?

Constant Co

- Laboratory procedures
  - Dilution
  - Spreading
  - Counting
  - Confirmation

**ISO 7218** 

Calculations

$$N = \frac{\sum C}{V \times 1, 1 \times d} = \frac{168 + 14}{1 \times 1, 1 \times 10^{-2}} = \frac{182}{0,011} =$$

Reporting

4.2 log cfu/g Campylobacter spp.

Final results



#### Results EURL-Campylobacter Proficiency Test Number 23 2019

Name of contact person (NRL)

Enumeration of Campylobacter in chicken meat

	Score	Performance
Overall enumeration	80.0%	Acceptable
Sensitivity identification (voluntary)	87.5%	Good

Analysis start

Country Testland

NRL lab ID

Laboratory The laboratory of food

100		Test Testsson			Test Testsson		3/12/2019		3/13/2019	
	Sample 1.	Sample 2.	Sample 3.	Sample 4.	Sample 5.	Sample 6.	Sample 7.	Sample 8.	Sample 9.	Sample 10.
Contents	Campylobacter jejuni	Campylobacter lari	Negative	Escherichia coli	Campylobacter lari	Campylobacter jejuni	Campylobacter jejuni Escherichia coli	Campylobacter coli	Campylobacter jejuni	Campylobacter coli
Batch No.	SLV306	SLV248	SLV289	SLV150	SLV299	SLV306	SLV313	SLV287	SLV305	SLV287

Name of contact person (PT 23)

Date of arrival

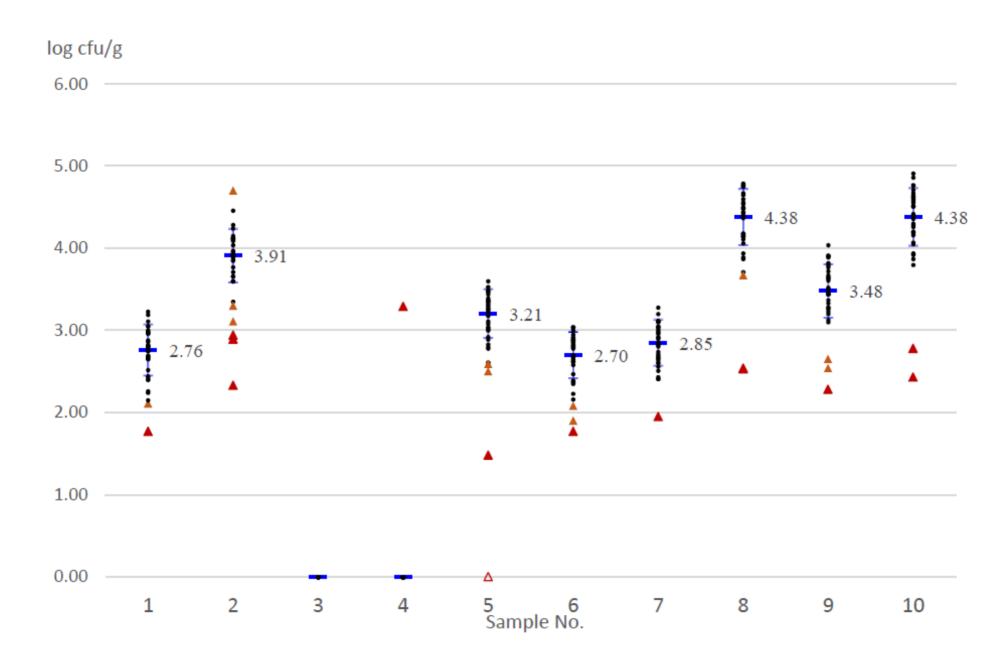
#### Enumeration of Campylobacter spp. (mandatory)

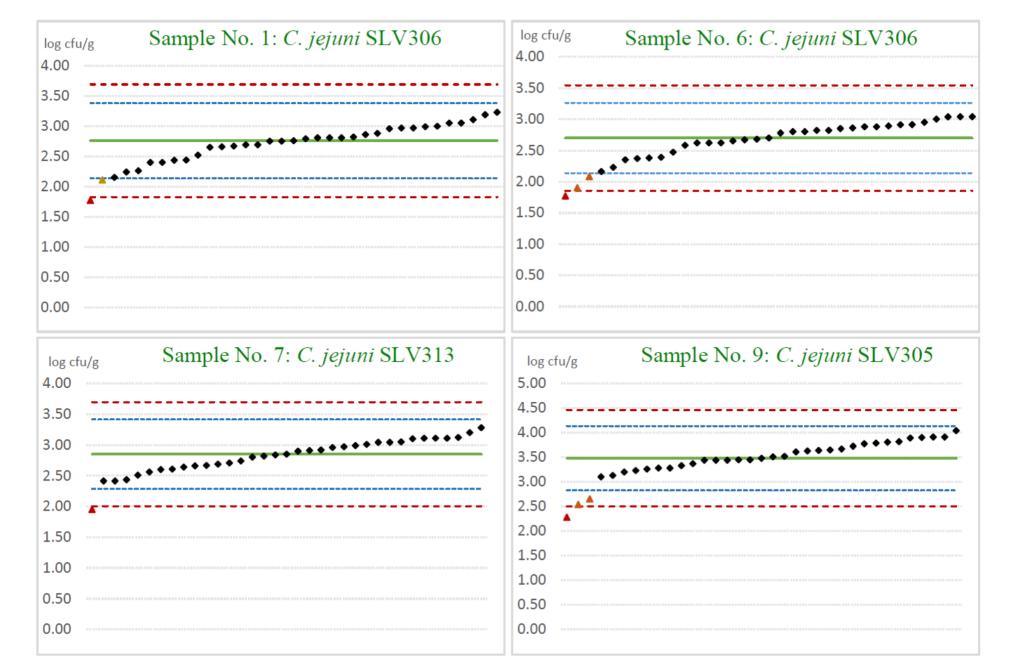
		above median value	–3σMAD / z-score +3σMAD / z-score			value between median value -2σMAD and -3σMAD / z-score between -2 and -3 value between median value +2σMAD and +3σMAD /z-score between 2 and 3				
Lab's results enumeration (log cfu/g)	2.75	4.45	<1.00	<1.00	2.56	2.58	2.69	2.68	4.43	4.56
Results as reported	2.75	4.45	0	<1.0	2.56	2.58	2.69	2.68	4.43	4.56
Score (points)	2	2	2	2	1	2	2	0	1	2
Z-score	-0.03	1.66	-	_	-2.18	-0.43	-0.57	-4.99	2.91	0.51
Median MADe oMADe	2.76 0.21 0.31	3.91 0.22 0.33	<1.00 - -	<1.00 - -	3.21 0.20 0.30	2.70 0.19 0.28	2.85 0.19 0.28	4.38 0.23 0.34	3.48 0.22 0.33	4.38 0.24 0.35
+2oMADe -2oMADe +3oMADe -3oMADe	3.39 2.13 3.70 1.82	4.57 3.25 4.89 2.93	- - -	- - -	3.80 2.61 4.10 2.31	3.27 2.13 3.55 1.85	3.42 2.28 3.70 2.00	5.07 3.69 5.41 3.35	4.14 2.82 4.46 2.50	5.09 3.67 5.44 3.32

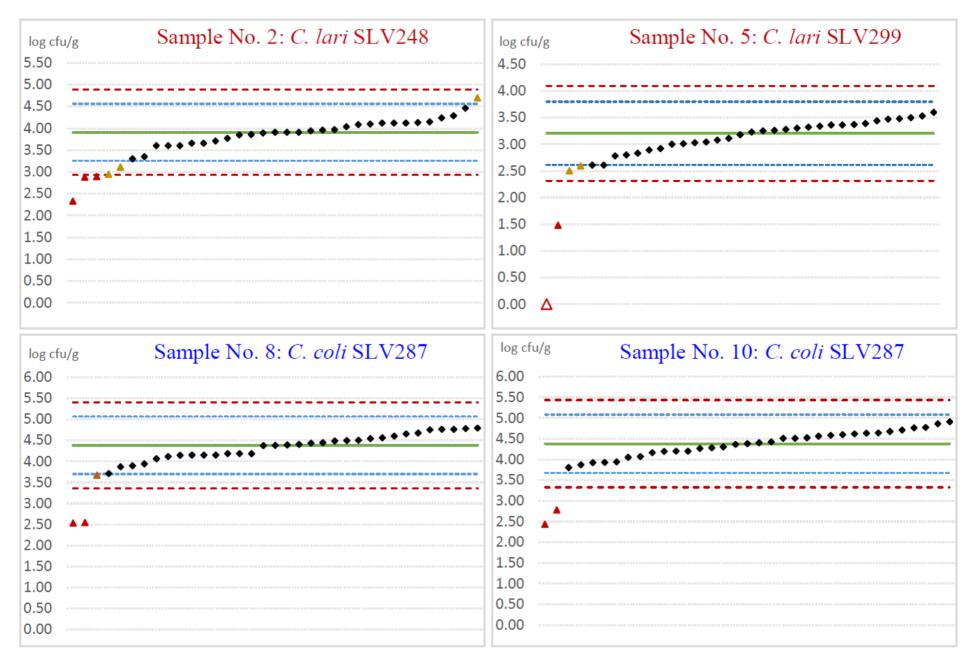
#### Species identification of Campylobacter spp. (voluntary)

incorrect/no species identification	false positive

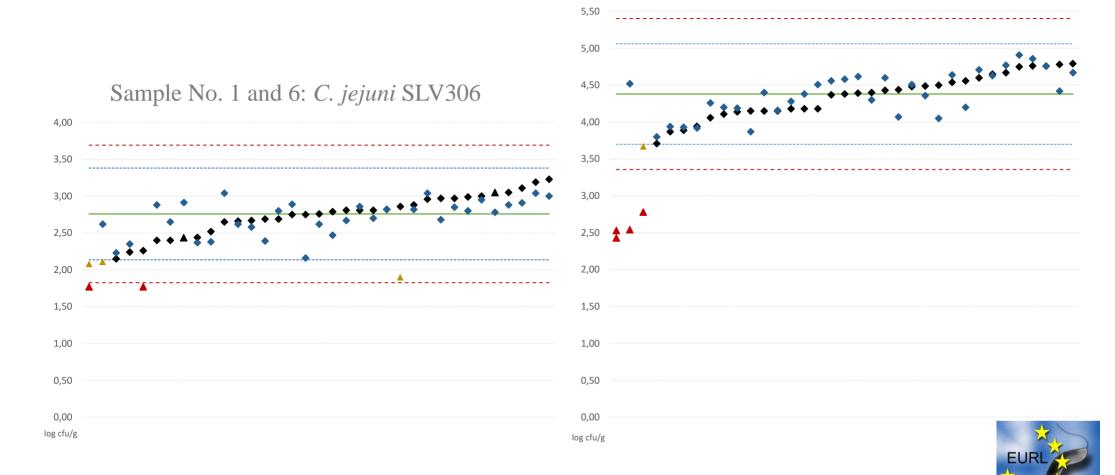
Lab's results species id entification	Campylobacter jejuni	Campylobacter lari	No growth at all	Growth of other, not Campylobacter	Campylobacter lari	Campylobacter coli	Campylobacter jejuni	Campylobacter coli	Campylobacter jejuni	Campylobacter coli
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Sample No. 8 and 10: C. coli SLV287



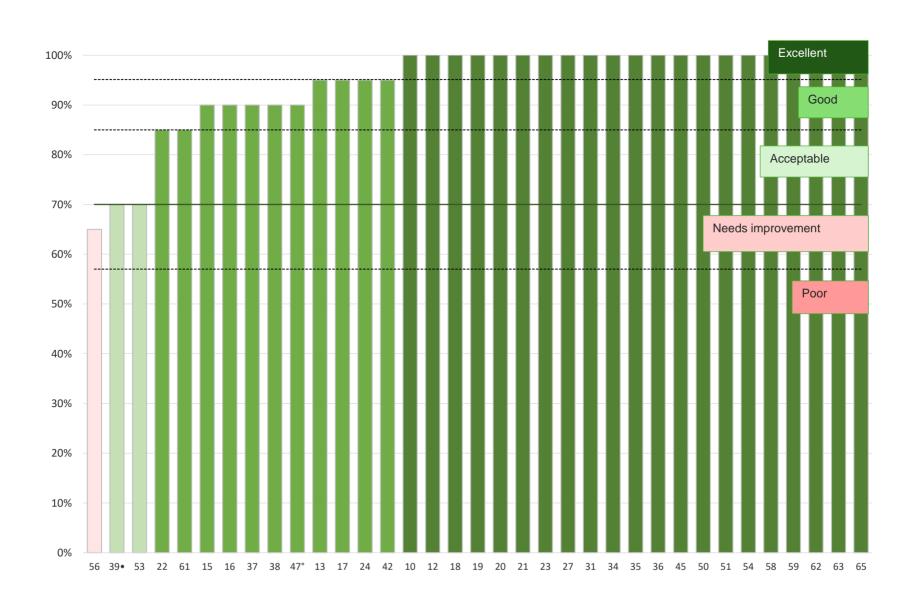
6.00

#### **HOW WAS PERFORMANCE CALCULATED?**

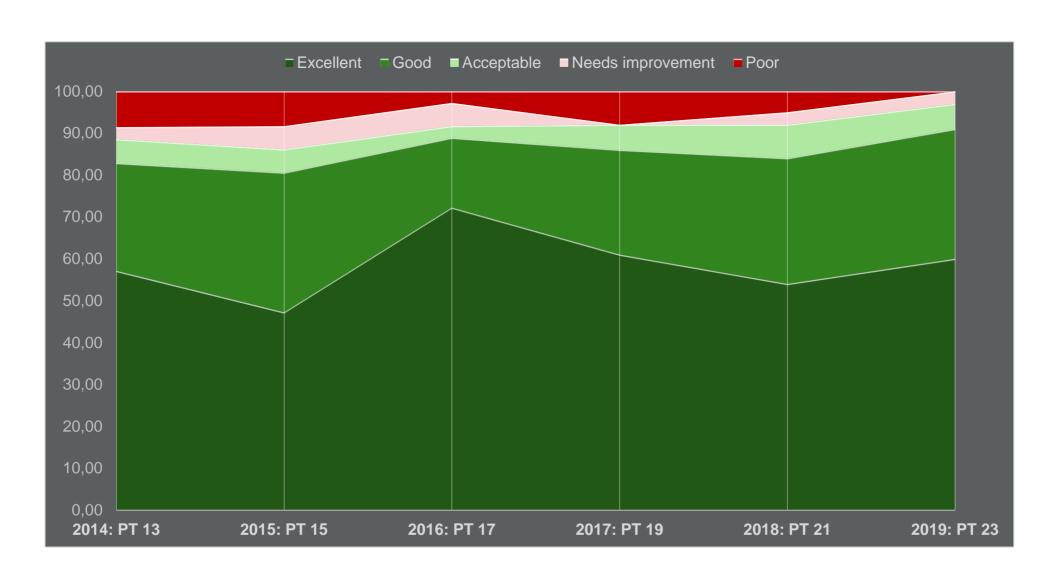
- The Median Absolute Deviation (MADe) to calculate performance
- $\sigma$ MADe = MADe × 1.4826
- Campylobacter-containing samples
  - Results within participants' median  $\pm 2\sigma MADe = 2$  points
  - Results between  $\pm 2\sigma MADe$  and  $\pm 3\sigma MADe = 1$  point
  - Results outside  $\pm 3\sigma MADe = 0$  points
- Campylobacter-negative samples
  - No Campylobacter reported = 2 points
  - False positive result = 0 points
- The maximum score (2 points for each sample) was 20 points
- Calculate the score for each participant

Grade	Scoring limits				
Excellent	20	95.1–100%			
Good	17–19	85.0–95.0%			
Acceptable	14–16	70.0–84.9%			
Needs improvement	12–13	57.0–69.9%			
Poor	<12	<57.0%			

### **PERFORMANCE PT 23**



# PERFORMANCE IN ENUMERATION OVER TIME



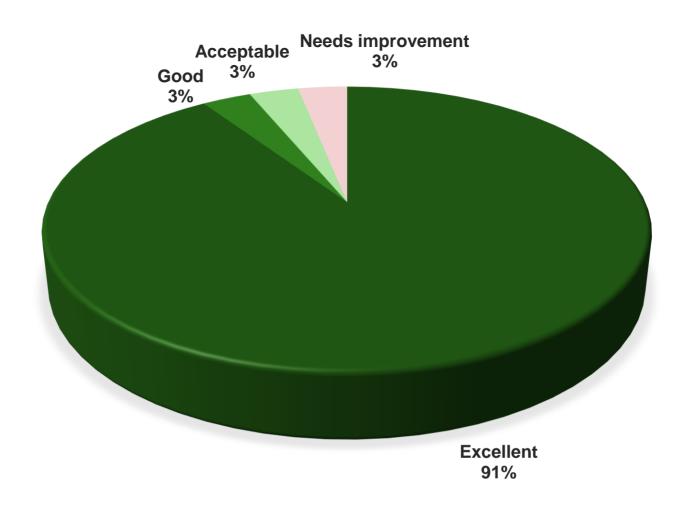
# PT 23: PERFORMANCE IN RELATION TO START OF ANALYSIS

Davi	Number			Performance	mance		
Day	of NRLs	Excellent Good A		Acceptable	Needs improvement	Poor	
12 <sup>th</sup> of March	2	1	1				
13 <sup>th</sup> of March	11	8	3				
14 <sup>th</sup> of March	3	2	1				
17 <sup>th</sup> of March	1		1				
18th of March	8	5	2	1			
19 <sup>th</sup> of March	3	1	1	1			
20th of March	1				1		
21st of March	1	1					
25st of March	4	2	2				
26 <sup>th</sup> of March	1	1					

## PT 23: SPECIES IDENTIFICATION (VOLUNTARY)

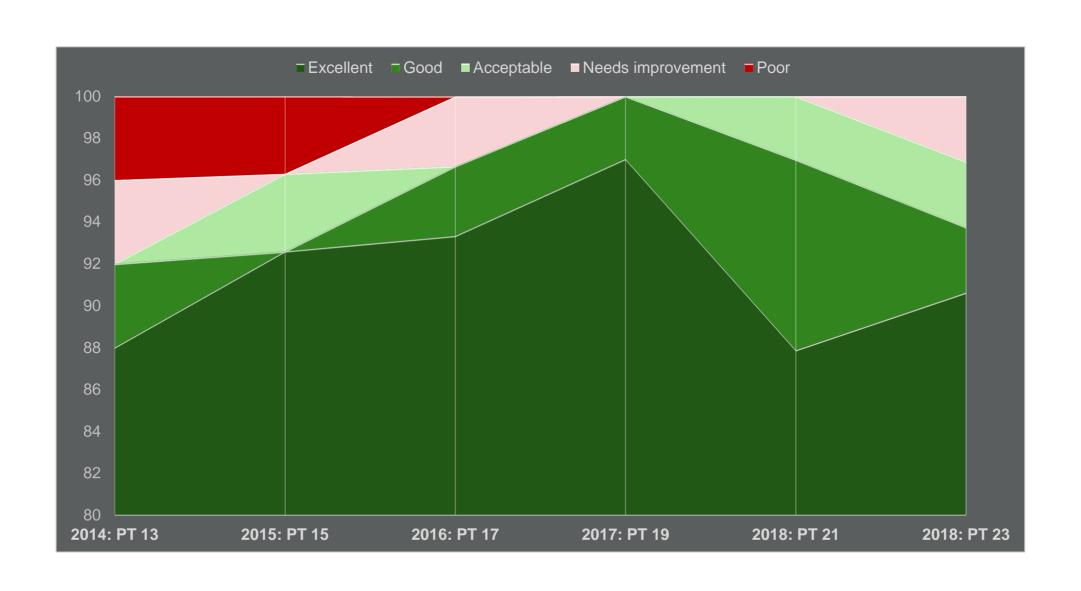
Content of sample (vial)	C. jejuni	C. coli	C. lari	Camp spp.	Other / No growth
1. <i>C. jejuni</i>	32				
2. <b>C. lari</b>			32		
3. Negative					32
4. E. coli					32
5. <b>C. lari</b>			31	1	
6. <i>C. jejuni</i>	32				
7. <b>C. jejuni</b> + E. coli	30		1	1	
8. <b>C.</b> coli	1	31			
9. <i>C. jejuni</i>	30	1		1	
10. <b>C.</b> coli		32			

# PERFORMANCE PT 23: SENSITIVITY IN SPECIES IDENTIFICATION (VOLUNTARY)





### PERFORMANCE IN IDENTIFICATION (SE) OVER TIME



# PT 24 – DETECTION AND SPECIES IDENTIFICATION OF CAMPYLOBACTER



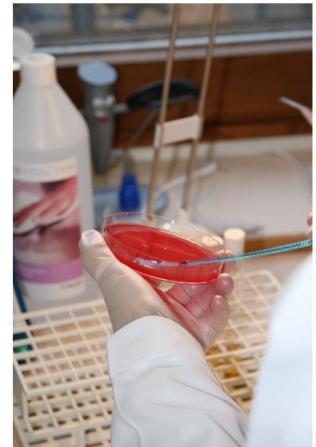
### **PROFICIENCY TEST NO. 24**

The objective was to assess the performance of the NRLs to detect and identify *Campylobacter* species in minced chicken meat and/or boot sock samples.

- Detection of Campylobacter spp. in minced chicken meat / boot sock samples
- Species identification of Campylobacter
- Three sets:
  - 10 core samples of minced chicken meat
  - 10 core sock samples mimicking samples taken in a chicken house with birds kept indoors
  - 4 educational sock samples (not included in the performance evaluation) mimicking samples taken in a chicken pen with birds kept outdoors
- Recommended method ISO 10272:2017, but other methods allowed
- Sample preparation boot socks: according to routine procedure in the laboratory or instructions sent out (for procedures A, B or C in ISO 10272:2017)

# PT 24: CONTENTS AND PROCEDURE: MINCED CHICKEN MEAT

- Plastic bag with minced chicken meat (about 120 g)
- 10 freeze-dried vials (with or without *Campylobacter*)
- Homogenise the meat (like a laboratory sample) before divided into 10 portions of 10 g
- From here on, treat each 10 g test portion as a separate sample
- Mix each vial with 10 g minced chicken meat
- Follow the method(s) of choice for
  - detection
  - species identification



of Campylobacter spp.



# PT 24: CONTENTS AND PROCEDURE: SOCK SAMPLES

- 10 numbered boot sock samples in plastic bags
- 10 freeze-dried vials (with or without Campylobacter)
- Mix each vial with enrichment broth or other liquid and pour into the bag with sock sample
- Follow the method(s) of choice for
  - detection
  - species identification







#### PT 24: CORE SAMPLES MINCED CHICKEN MEAT

Sample No.	Content in vial	Batch No.	Level	log cfu/vial
11	Campylobacter lari	SVA016	High	4.38
12	Campylobacter jejuni	SVA021	High	4.28
13	Campylobacter coli	SVA023	Low	2.93
14	Negative	SLV289		
15	Negative	SLV289		
16	Campylobacter coli	SVA022	High	3.45
17	Campylobacter lari	SVA017	Low	3.27
18	Campylobacter jejuni	SVA027	Low	2.02
19	Campylobacter jejuni	SVA025	Low	3.20
20	Campylobacter jejuni	SVA025	Low	3.20

Candida albicans was added as background flora in the chicken meat







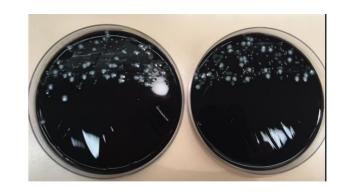
#### PT 24: BOOT SOCK SAMPLES

Sample No.	Content in vial	Batch No.	Level	log cfu/vial	Sock
21	Campylobacter jejuni	SVA025	Low	3.20	E. coli
22	Negative	SLV289			E. coli
23	Campylobacter jejuni	SVA027	Low	2.02	
24	Campylobacter jejuni	SVA021	High	4.28	E. coli
25	Campylobacter lari	SVA017	Low	3.27	
26	Campylobacter jejuni	SVA025	Low	3.20	
27	Campylobacter lari	SVA016	High	4.38	E. coli
28	Campylobacter coli	SVA022	High	3.45	
29	Negative	SLV289			
30	Campylobacter coli	SVA023	Low	2.93	E. coli

#### **EDUCATIONAL SAMPLES**

Sample No.	Content in vial	Batch No.	Level (log cfu/vial)
31	Campylobacter lanienae	SVA019	3.75
32	Campylobacter helveticus	SVA026	6.10
33	Campylobacter upsaliensis	SVA018	4.47
34	Campylobacter lari + Campylobacter jejuni	SVA015	4.48 (in total)

## PT 24: QUALITY CONTROL



- Vials produced by EURL or the National Food Agency (negatives)
- Tested for homogeneity and stability by the producer
- Non-Campylobacter (E. coli, Candida albicans) strains were tested for use as live cultures
- Vials together with matrix were analysed according to ISO 10272-1:2017:
  - Chicken meat: procedure A (Bolton)
  - Sock samples: procedure A (Bolton), B (Preston) and C (direct streaking)
- Tested three times, once before and twice after dispatch





# PT 24: PREPARATION OF THE MATRIX: MINCED CHICKEN MEAT

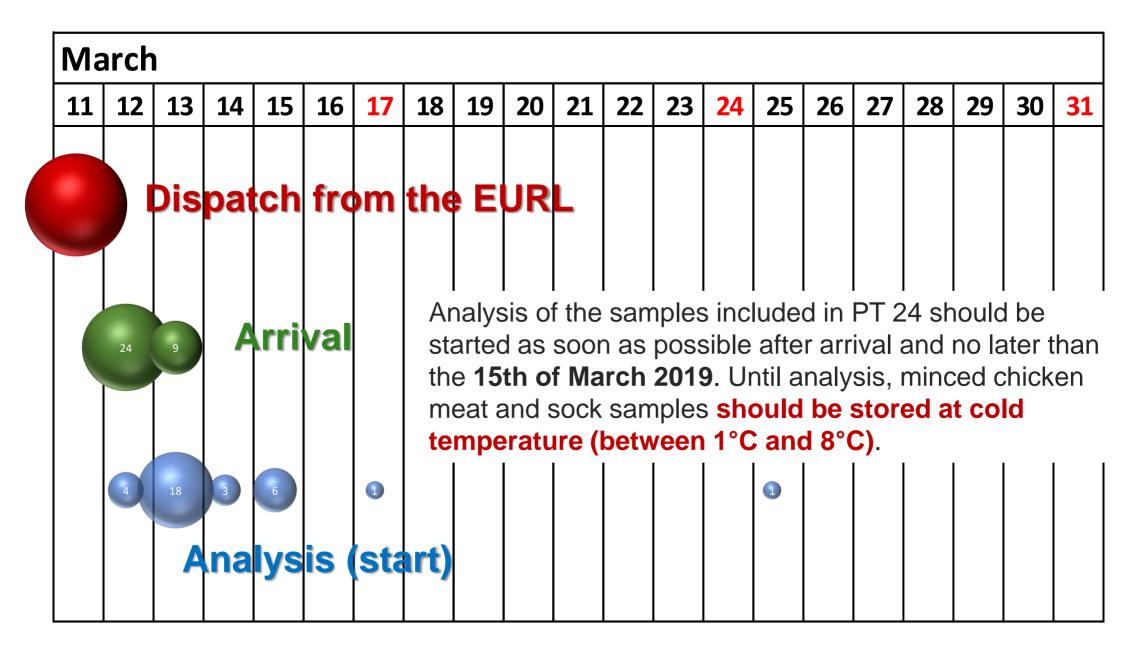
- Campylobacter-free chicken meat was grinded and refreezed
- Minced chicken meat was thawed at 4 °C
- An overnight culture with Candida albicans was prepared
- On the day of dispatch, the minced chicken meat was
- mixed with theovernight culture, homogenised and divided in 120 g portions (one for each participant)

# PT 24: PREPARATION OF THE MATRIX: BOOT SOCK SAMPLES

- An overnight culture with E. coli was prepared
- Campylobacter-free caeca were cut and placed in a stomacher bag and mixed with Cary Blair transport medium
- For samples with background, the overnight culture was mixed with the caecum suspension
- For samples without background, serum broth of the same volume was added to the caecum suspension
- 20 ml of the suspension (with or without background) were added to a plastic bag with a boot sock, one for each sample
- The sock samples were stored at 4 °C over the weekend



#### PT 24: TIME TO ARRIVAL & START OF ANALYSIS



### PT 24: METHODS AND PROCEDURES

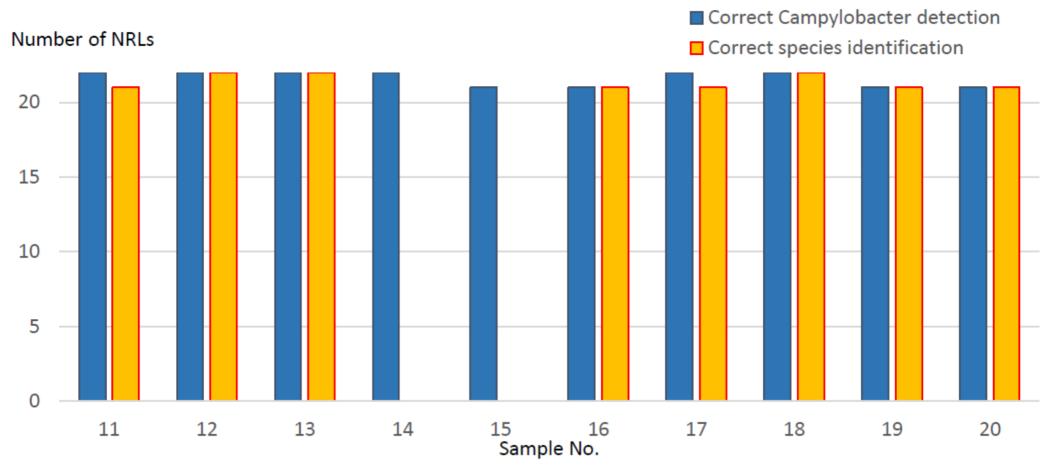
Test	ISO 10272- 1:2017	Other methods	Enrich -ment	Bolton (A)	Preston (B)	Other	Direct (C)
Meat	19	3	22	16	7	1	2
Socks	21	1	20	10	10		5
Educ	23	4	24	15	10	1	9





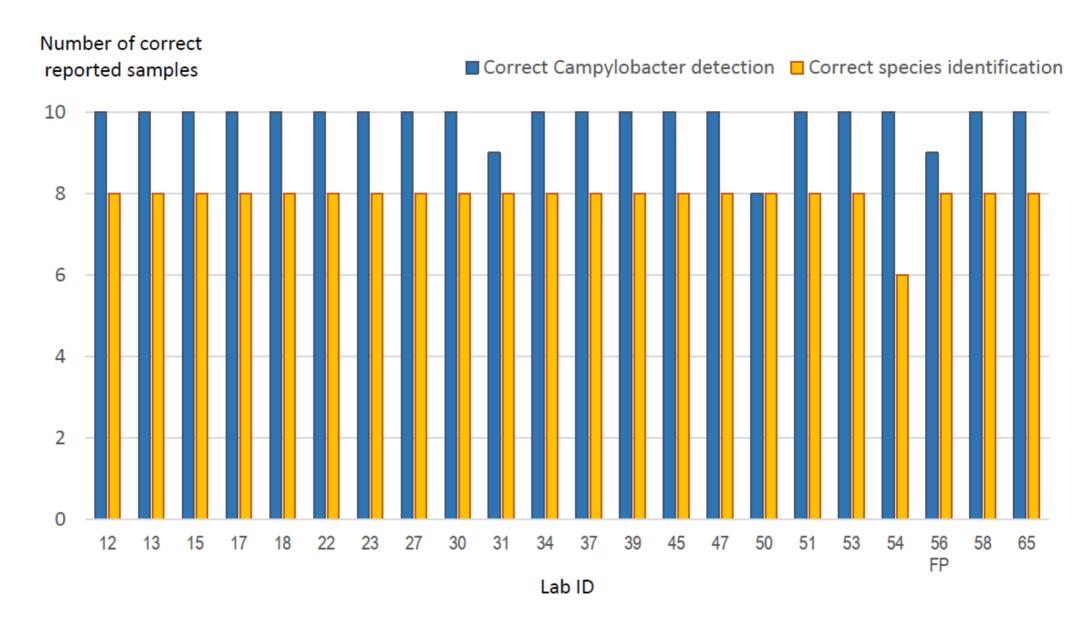


## PT 24: CORRECT REPORTED RESULTS PER SAMPLE (MINCED CHICKEN MEAT)

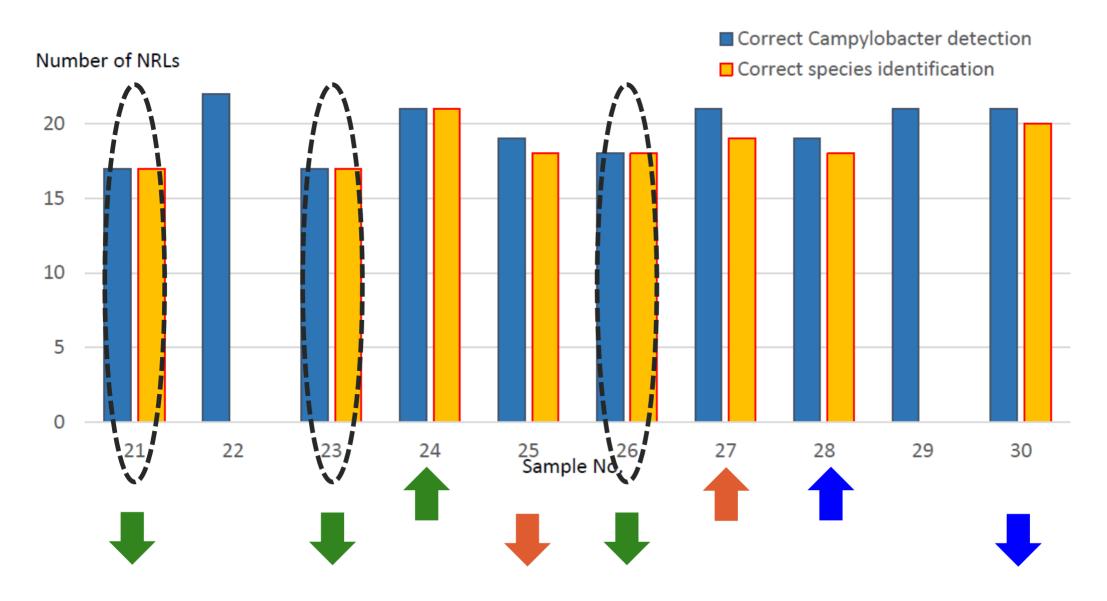




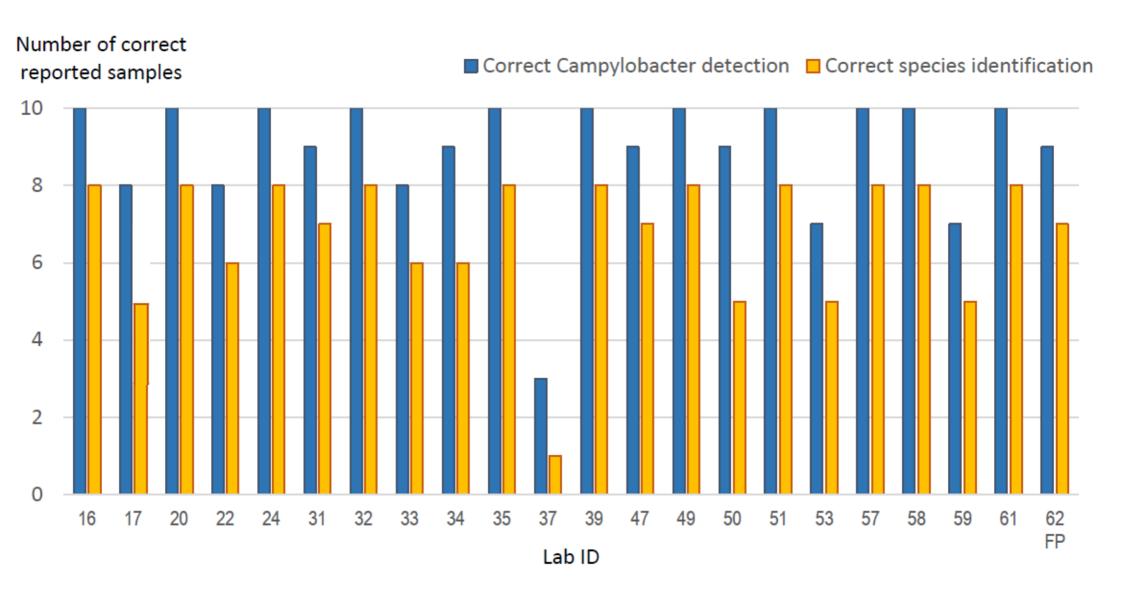
# PT 24: CORRECT REPORTED RESULTS PER LAB (MINCED CHICKEN MEAT)



## PT 24: CORRECT REPORTED RESULTS PER SAMPLE (SOCK SAMPLES)

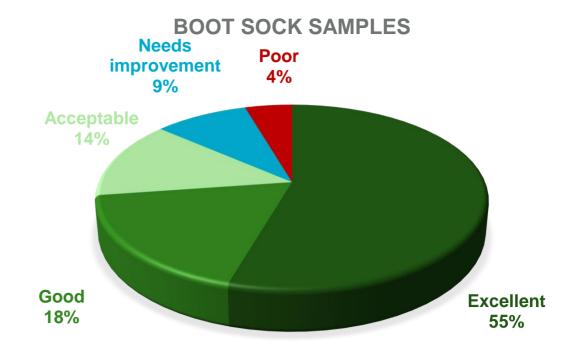


# PT 24: CORRECT REPORTED RESULTS PER LAB (SOCK SAMPLES)



### PT 24: PERFORMANCE – SENSITIVITY (SE) IN DETECTION OF CAMPYLOBACTER





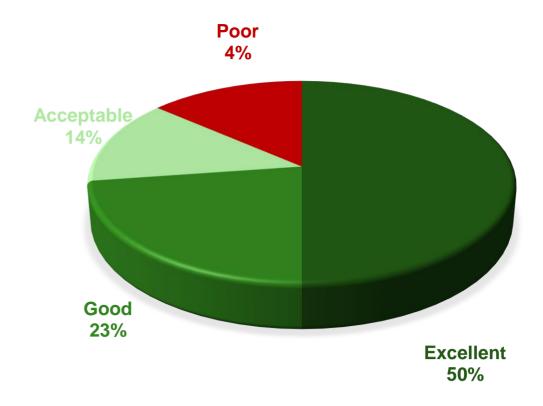


## PT 24: ACCURACY IN DETECTING POSITIVE AND NEGATIVE CAMPYLOBACTER SAMPLES

#### MINCED CHICKEN MEAT

# Acceptable Good 9% Excellent 86%

#### **BOOT SOCK SAMPLES**



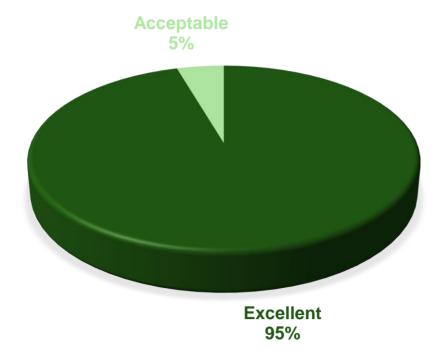


#### PT 24: REPORTED SPECIES IDENTIFICATION

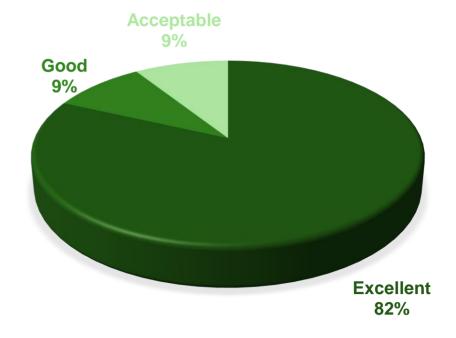
Sample No.	Bacterial species	C. jejuni	C. coli	C. lari	Campylobacter spp. but unable to identify species	Growth of other, not <i>Campylobacter</i>	No growth at all
11 / 27	Campylobacter lari	0+1	0+1	21+19	1+0	0+1	
12 / 24	Campylobacter jejuni	22+21					0+1
13 / 30	Campylobacter coli		22+20	0+1			
14 / 22	Negative					7+18	15+4
15 / 29	Negative	0+1	1+0			8+15	13+6
16 / 28	Campylobacter coli		21+18	0+1		0+2	0+1
17 / 25	Campylobacter lari		0+1	21+18	1+0	0+2	0+1
18 / 23	Campylobacter jejuni	22+17				0+3	0+2
19 / 21	Campylobacter jejuni	21+17				1+3	0+2
20 / 26	Campylobacter jejuni	21+18				1+2	0+2

## PT 24: PERFORMANCE – SENSITIVITY SPECIES IDENTIFICATION

#### MINCED CHICKEN MEAT



#### **BOOT SOCK SAMPLES**





#### PT 24: EDUCATIONAL SAMPLES

Sample No.	Bacterial species	C. jejuni	Both C. jejuni and C. lari	C. lari	C. upsaliensis	C. helveticus	C. lanienae	C. cuniculorum	Campylobacter spp. but unable to identify species	No Campylobacter detected
31	Campylobacter lanienae	1		1			11		7	6
32	Campylobacter helveticus					9		1		17
33	Campylobacter upsaliensis				17				1	9
34	Campylobacter lari Campylobacter jejuni	11	11	4					1	

## PT 24: OVERALL SENSITIVITY AND PERFORMANCE RATE FOR EDUCATIONAL SAMPLES

Sample No.	Campylobacter species	Sensitivity in detection	Sensitivity in species id	Combined performance rate
31	C. lanienae	77.8%	52.4%	59.3%
32	C. helveticus	37.0%	90.0%	35.2%
33	C. upsaliensis	66.7%	94.4%	64.8%
34	C. lari + C. jejuni	100.0%	68.5%	84.3%
All		70.4%	73.0%	60.9%



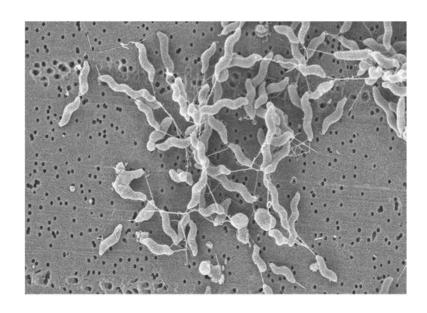
#### **COMMENTS AND QUESTIONS**

- Sock samples:
  - How much liquid should be used to one sock sample?
  - Which procedure (A, B, C) is most adequate?









#### **THANK YOU!**



